## Amendments to the Claims:

This listing of claims will replace all prior versions, and listings of claims in the application:

## **Listing of Claims:**

## 1-83 (canceled)

1 84 (previously presented): A material having a fluorogenic moiety linked to a 2 solid support, said material having the structure:

wherein:

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4 R<sup>1</sup>, R<sup>3</sup>, R<sup>4</sup> and R<sup>6</sup> are each H; 5 R<sup>2</sup> is -NHR<sup>15</sup>; and 6 R<sup>5</sup> is -R<sup>14</sup>-SS, 7 wherein:

 $R^{14}$  is  $-CH_2C(O)NH-$ ;

R<sup>15</sup> is a member selected from the group consisting of amine protecting groups, -C(O)-AA and -C(O)-P:

11 wherein:

P is a peptide sequence;

13 AA is an amino acid residue; and

SS is a solid support.

- 85 (previously presented): The material in accordance with claim 84, wherein
   R<sup>15</sup> is an amine protecting group.
- 1 86 (previously presented): The material in accordance with claim 85, wherein 2 said amine protecting group is 9-fluorenylmethoxycarbonyl (Fmoc).
- 87 (previously presented): The material in accordance with claim 84, wherein
   R<sup>15</sup> is -C(O)-AA, wherein AA is an amino acid residue.
- 88 (previously presented): The material in accordance with claim 84, wherein
   R<sup>15</sup> is -C(O)-P, wherein P is a peptide sequence.
- 1 89 (previously presented): The material in accordance with claim 84, wherein the solid support is a Rink resin.
- 90 (previously presented): A material having a fluorogenic moiety linked to a
   solid support, said material having the structure:

4 wherein:

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- 5 SS is a solid support, wherein said the support is a Rink resin.
- 91 (previously presented): A library of fluorogenic peptides comprising sublibraries P1, P2, P3 and P4, wherein each of the sub-libraries P1, P2, P3 and P4 comprises tetrapeptides having the structure:

5 wherein:

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6 SS is a solid support, and

7 wherein:

8 for sub-library P1, each AA<sup>1</sup> is a different amino acid of the 20 amino acids, and 9 each of AA<sup>2</sup>-AA<sup>4</sup> is an isokinetic mixture of 20 amino acids:

for sub-library P2, each of  $AA^2$  is a different amino acid of the 20 amino acids, and each of  $AA^1$ ,  $AA^3$  and  $AA^4$  is an isokinetic mixture of 20 amino acids;

for sub-library P3, each of AA<sup>3</sup> is a different amino acid of the 20 amino acids, and each of AA<sup>1</sup>, AA<sup>2</sup> and AA<sup>4</sup> is an isokinetic mixture of 20 amino acids; and

for sub-library P4, each of AA<sup>4</sup> is a different amino acid of the 20 amino acids, and each of AA<sup>1</sup>, AA<sup>2</sup> and AA<sup>4</sup> is an isokinetic mixture of 20 amino acids.

1 92 (previously presented): The library in accordance with claim 91, wherein the 20 amino acids are the 20 naturally occurring amino acids excluding cysteine and including norleucine.

1 93 (previously presented): The library in accordance with claim 91, wherein the solid support is a Rink resin.

94 (previously presented): A library of fluorogenic peptides comprising sublibraries P1, P2, P3 and P4, wherein each of the sub-libraries P1, P2, P3 and P4 comprises tetrapeptides having the structure:

5 wherein:

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for sub-library P1, each AA<sup>1</sup> is a different amino acid of the 20 amino acids, and
each of AA<sup>2</sup>-AA<sup>4</sup> is an isokinetic mixture of 20 amino acids;

for sub-library P2, each of AA<sup>2</sup> is a different amino acid of the 20 amino acids, and each of AA<sup>1</sup>, AA<sup>3</sup> and AA<sup>4</sup> is an isokinetic mixture of 20 amino acids;

for sub-library P3, each of AA<sup>3</sup> is a different amino acid of the 20 amino acids.

and each of AA<sup>1</sup>, AA<sup>2</sup> and AA<sup>4</sup> is an isokinetic mixture of 20 amino acids; and
for sub-library P4, each of AA<sup>4</sup> is a different amino acid of the 20 amino acids,
and each of AA<sup>1</sup>, AA<sup>2</sup> and AA<sup>4</sup> is an isokinetic mixture of 20 amino acids.

1 95 (previously presented): The library in accordance with claim 94, wherein the 20 amino acids are the 20 naturally occurring amino acids excluding cysteine and including norleucine.

96 (previously presented): A method of determining a peptide sequence
 specificity profile of an enzymatically active protease, said method comprising:

- (a) contacting said protease with a library of peptides according to claim 91 or
   claim 94 in such a manner whereby the fluorogenic moiety is released
   from the peptide sequence, thereby forming a fluorescent moiety;
- (b) detecting said fluorescent moiety;
- (c) determining the sequence of said peptide sequence, thereby determining said
   peptide sequence specificity profile of said protease.
- 97 (previously presented): The method according to claim 96, further comprising
   (d) quantifying said fluorescent moiety, thereby quantifying said protease.
- 1 98 (previously presented): The method according to claim 97, wherein said 2 protease is a member selected from the group consisting of aspartic protease, cysteine protease, 3 metalloprotease and serine protease.
- 99 (previously presented): A library of fluorogenic peptides comprising sub libraries P1, P2, P3 and P4, wherein each of the sub-libraries P1, P2, P3 and P4 comprises
   hexapeptides having the structure:

- 5 wherein:
- 6 SS is a solid support, and
- 7 wherein:

8 for each sub-library P1, P2, P3 and P4, AA<sup>1</sup>, AA<sup>2</sup>, AA<sup>3</sup> and AA<sup>4</sup> in each of the 9 hexapeptides are the same amino acid residues;

for sub-library P1, each of AA<sup>5</sup> is a different amino acid of the 20 amino acids,
and each of AA<sup>6</sup>, AA<sup>7</sup> and AA<sup>8</sup> is an isokinetic mixture of 20 amino acids;

12 for sub-library P2, each of AA<sup>6</sup> is a different amino acid of the 20 amino acids,

13 and each of  $AA^5$ ,  $AA^7$  and  $AA^8$  is an isokinetic mixture of 20 amino acids;

for sub-library P3, each of AA7 is a different amino acid of the 20 amino acids,

15 and each of AA5, AA6 and AA8 is an isokinetic mixture of 20 amino acids; and

for sub-library P4, each of AA<sup>8</sup> is a different amino acid of the 20 amino acids,
 and each of AA<sup>5</sup>, AA<sup>6</sup> and AA<sup>7</sup> is an isokinetic mixture of 20 amino acids.

1 100 (previously presented): The library in accordance with claim 99, wherein the 20 amino acids are the 20 naturally occurring amino acids excluding cysteine and including norleucine.

1 101 (previously presented): The library in accordance with claim 99, wherein the 2 solid support is a Rink resin.

1 102 (previously presented): A library of fluorogenic peptides comprising sublibraries P1, P2, P3 and P4, wherein each of the sub-libraries P1, P2, P3 and P4 comprises hexapeptides having the structure:

5 wherein

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6	for each sub-library P1, P2, P3 and P4, AA1, AA2, AA3 and AA4 in each of the
7	hexapeptides are the same amino acid residues;
8	for sub-library P1, each of AA5 is a different amino acid of the 20 amino acids,
9	and each of AA <sup>6</sup> , AA <sup>7</sup> and AA <sup>8</sup> is an isokinetic mixture of 20 amino acids;
10	for sub-library P2, each of AA <sup>6</sup> is a different amino acid of the 20 amino acids,
11	and each of AA <sup>5</sup> , AA <sup>7</sup> and AA <sup>8</sup> is an isokinetic mixture of 20 amino acids;
12	for sub-library P3, each of AA7 is a different amino acid of the 20 amino acids,
13	and each of AA5, AA6 and AA8 is an isokinetic mixture of 20 amino acids; and
14	for sub-library P4, each of AA8 is a different amino acid of the 20 amino acids,
15	and each of AA <sup>5</sup> , AA <sup>6</sup> and AA <sup>7</sup> is an isokinetic mixture of 20 amino acids.
1	103 (previously presented): The library in accordance with claim 102, wherein
2	the 20 amino acids are the 20 naturally occurring amino acids excluding cysteine and including
3	norleucine.
1	104 (previously presented): A method of determining a peptide sequence
2	specificity profile of an enzymatically active protease, said method comprising:
3	(a) contacting said protease with a library of peptides according to claim 99 or
4	claim 102 in such a manner whereby the fluorogenic moiety is released
5	from the peptide sequence, thereby forming a fluorescent moiety;
6	(b) detecting said fluorescent moiety;
7	(c) determining the sequence of said peptide sequence, thereby determining said
8	peptide sequence specificity profile of said protease.
1	105 (previously presented): The method according to claim 104, further

comprising (d) quantifying said fluorescent moiety, thereby quantifying said protease.

1 106 (previously presented): The method according to claim 105, wherein said 2 protease is a member selected from the group consisting of aspartic protease, cysteine protease, 3 metalloprotease and serine protease.

107 (previously presented): A library of twenty fluorogenic amino acid amides
 having the structure:

4 wherein:

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SS is a solid support, and

each AA<sup>1</sup> for the twenty fluorogenic amino acid amides is a different amino acid
 residue.

1 108 (previously presented): The library in accordance with claim 107, wherein
2 the amino acid residues are the 20 naturally occurring amino acids excluding cysteine and
3 including norleucine.

1 109 (previously presented): The library in accordance with claim 108, wherein 2 the solid support is a Rink resin.

1 110 (previously presented): A library of twenty fluorogenic amino acids having 2 the structure:

3 4 wherein:

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each AA1 for the twenty fluorogenic amino acids is a different amino acid residue

- 1 111 (previously presented): The library in accordance with claim 110, wherein
  the amino acid residues are the 20 naturally occurring amino acids excluding cysteine and
  including norleucine..
- 1 112 (previously presented): A method of determining an amino acid specificity
  2 profile of an enzymatically active protease, said method comprising:
  - (a) contacting said protease with a library of amino acids according to claim 108
    or claim 110 in such a manner whereby the fluorogenic moiety is released
    from the amino acid, thereby forming a fluorescent moiety;
- (b) detecting said fluorescent moiety;
  - (c) determining the identity of the amino acid, thereby determining said amino acid specificity profile of said protease.
- 1 113 (previously presented): The method according to claim 112, further comprising (d) quantifying said fluorescent moiety, thereby quantifying said protease.
- 1 114 (previously presented): The method according to claim 113, wherein said 2 protease is a member selected from the group consisting of aspartic protease, cysteine protease, 3 metalloprotease and serine protease.